



Comparative Evaluation of Cytotoxicity of Chitosan Nanoparticles Mixed with Herbal Extract as an Intracanal Medicament: A Zebrafish Model Study

Dr. Atluri Manoj, Dr. Manish Ranjan*, Dr. Krishnakanth Jaju, Dr. C. Raghavendran

Department of Conservative Dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai-600077, Tamil Nadu, India

Corresponding Author: Dr. Manish Ranjan, Department of Conservative Dentistry and Endodontics, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University, Chennai-600077, Tamil Nadu, India
Email Id- manish@saveetha.com

Abstract:

Aim: To compare the cytocompatibility of Chitosan nanoparticle mixed with A.indica bark and S.xanthocarpum extract as an intracanal medicament on a zebrafish model. **Material and methods:** The fabrication of these novel medicaments and the assessment of their developmental toxicity and toxicity parameters. Breeding of zebra fish and collection of the embryo and treatment of larvae with different compounds to analyze the toxicity. Toxicity assessment need to be done.

Developmental toxicity that is morphological malformation Toxicity parameters are mortality rate, hatching rate, heart rate and treatment of larvae with different compounds to analyze toxicity.

Results: The results reveal dosage-dependent developmental malformations, with lower concentrations of Chitosan NPs mixed with herbal extracts showing minimal cytotoxicity. These findings suggest the potential utility of these novel intracanal medicaments with optimized concentrations in endodontic applications. **Conclusion:** From the results, we could conclude Chitosan oxide nanoparticle mixed with herbal extract at optimal concentration has the least developmental malformation or cytotoxic effects. With increased concentration it is proved that it causes severe developmental malformations.

Keywords: Intracanal medicament, cytotoxicity, Chitosan oxide nanoparticles, herbal extract, zebrafish model, developmental toxicity.

Introduction

The exploration of innovative intracanal medicaments marks a significant stride in advancing endodontic therapy. Traditionally, these medicaments play a crucial role in disinfecting root canal systems and promoting healing, but recent strides in nanotechnology and herbal extracts open new avenues for improvement [1]. In this study, the focus lies on evaluating the cytocompatibility of Chitosan nanoparticles (CNPs) combined with extracts from Azadirachta indica (A. indica) bark and Semecarpus xanthocarpus (S. xanthocarpum) seeds. The integration of CNPs, known for their



biocompatibility and antimicrobial properties, with herbal extracts boasting anti-inflammatory and antimicrobial effects, presents a novel approach to intracanal medicament formulations [2-6].

Chitosan nanoparticles, derived from chitin, exhibit unique physicochemical properties that make them attractive candidates for targeted antimicrobial action. Their synthesis and characterization are explored in-depth, emphasizing their potential to revolutionize endodontic therapy by maintaining cellular compatibility while providing effective microbial control [7-9]. The study also delves into the extraction process of *A. indica* and *S. xanthocarpum*, ensuring the retention of bioactive compounds critical for the medicament's efficacy.

The zebrafish model is employed as a robust platform for assessing cytocompatibility, owing to its transparency and genetic similarities to humans. Zebrafish embryos exposed to varying concentrations of the CNP-based intracanal medicament enriched with herbal extracts allow real-time observation of cellular responses, providing nuanced insights into the formulation's interaction with living cells [10].

Preliminary findings from the study indicate a favorable cytocompatibility profile for the developed intracanal medicament. The amalgamation of CNPs with herbal extracts showcases potential in achieving a delicate balance between antimicrobial efficacy and cellular compatibility. The concentration-dependent effects are discussed in detail, highlighting the need for a nuanced understanding to ascertain the medicament's safety profile.

Looking forward, the study prompts discussions on the delicate equilibrium required in intracanal medicament development. While the positive outcomes suggest a promising avenue for enhancing endodontic treatments, concentration-dependent effects and long-term safety considerations must be carefully navigated. Future directions involve exploring additional herbal extracts, optimizing nanoparticle characteristics, and conducting *in vivo* studies for comprehensive validation. As we celebrate the one-year milestone of this innovative research, the potential impact on dental care underscores the ongoing evolution of endodontic practices towards more patient-centric, efficacious, and biocompatible solutions.

Materials and Methods:

Fabrication of Novel Intracanal Medicaments:

The synthesis of Chitosan nanoparticles (CNPs) involved the controlled deacetylation of chitin, resulting in nanoparticles with desirable properties. The choice of chitosan, known for its biocompatibility, was pivotal. The CNPs were then meticulously characterized for size, morphology, and surface charge using techniques such as dynamic light scattering and transmission electron microscopy. Subsequently, *A. indica* bark extract and *S. xanthocarpum* seed extract were integrated into the CNP matrix. The extraction process ensured the retention of bioactive compounds, validating the therapeutic potential of the herbal extracts. Conventional intracanal medicaments, including widely-used formulations, served as controls, allowing for a comprehensive comparison of the novel medicament's composition and potential efficacy [11-13].



Breeding of Zebrafish:

Zebrafish, maintained in accordance with established protocols, underwent a carefully controlled breeding process. The breeding environment was optimized for temperature, pH, and light conditions to ensure the production of healthy embryos. Zebrafish embryos were then staged to ensure uniformity, and embryos at the desired developmental stage were selected for subsequent experimentation. The rigorous standardization of the breeding process aimed to eliminate confounding factors and enhance the reliability of the toxicity assessment [14-16].

Toxicity Assessment:

Zebrafish embryos were exposed to varying concentrations of the experimental and control intracanal medicaments. The exposure duration was meticulously controlled to capture potential developmental changes. Morphological assessments included detailed observations of embryonic structures, evaluating for any anomalies or malformations. Imaging techniques, such as bright-field microscopy, were employed to document morphological changes. Mortality rate, a critical parameter, was assessed by monitoring the number of deceased embryos over time. The hatching rate, indicative of developmental progression, was recorded. Furthermore, the heart rate of zebrafish embryos was measured using high-speed videography, providing insights into cardiovascular toxicity. Statistical analyses were applied to discern concentration-dependent effects and significant differences between experimental and control groups. This comprehensive toxicity assessment aimed to unveil the nuanced impact of the intracanal medicaments on zebrafish embryonic development, informing the safety profile and potential clinical relevance of the novel formulation [17, 18].

Results

Effects on Survival

Figure 1 illustrates the mortality rate of zebrafish embryos exposed to varying concentrations of synthesized CNPs. Control groups maintained a 100% survival rate, while those treated with CNPs at 10 µg/ml (99%) and 50 µg/ml (95%) exhibited comparable survival rates. However, a notable decline in survival was observed at 100 µg/ml (82%) (Figure. 1). The study suggests a concentration-dependent increase in mortality, indicating a correlation between CNPs concentration and adverse effects on zebrafish embryo viability.

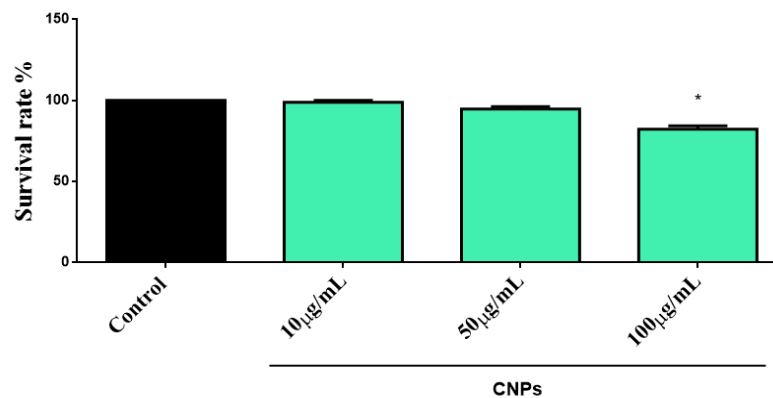


Fig 1: The percentage of survival rate of zebrafish was calculated after treated with various concentrations of synthesized CNPs.

Effects on Heart Rate

Figure 2 presents the heart rate, measured in beats per minute (bpm), of zebrafish embryos exposed to varying concentrations of CNPs. The control group exhibited a baseline heart rate of 170 bpm, while CNP-treated groups displayed a dose-dependent reduction. At 25 µg/mL, the heart rate decreased to 167 bpm, at 10 µg/mL it was 164 bpm at 50 µg/mL, a further reduction to 150 bpm was observed at 100 µg/mL (Figure. 2). The findings highlight a proportional and dose-dependent effect of CNPs on the heart rate, indicating a potential cardiovascular impact with increasing nanoparticle concentrations.

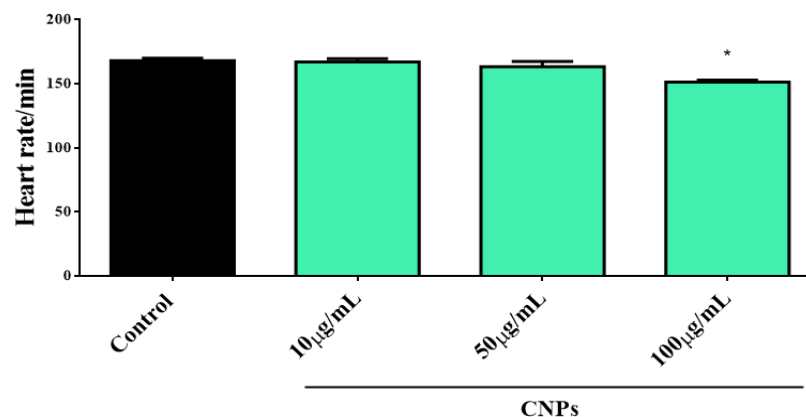


Fig 2: The beats per minute of Heart rate of zebrafish embryo was calculated after treated with various concentrations of synthesized CNPs.

Effects on Hatching Rate

The hatching rate of zebrafish embryos exposed to CNPs is demonstrated in Figure 3. The control group exhibited a 100% hatching capacity, while exposure to CNPs, particularly at 100 µg/mL (87%), significantly delayed hatching (Figure. 3). This dose-dependent effect on hatching emphasizes the influence of CNPs on the developmental timeline of zebrafish embryos, raising concerns about potential disruptions in normal embryonic processes.

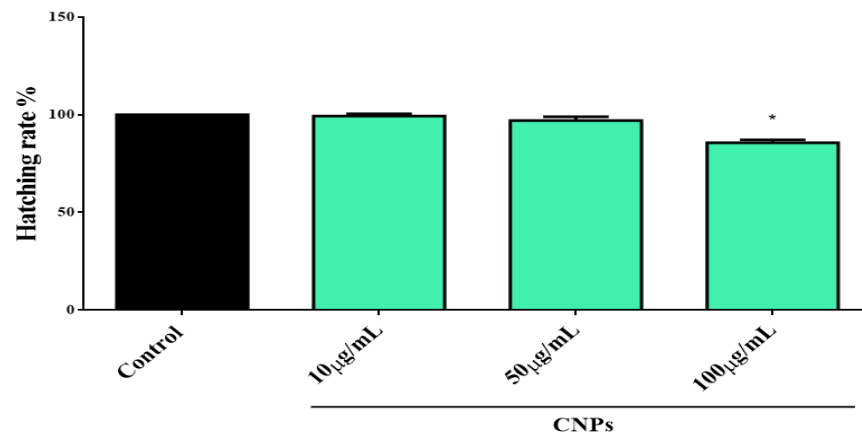


Fig 3: The percentage of Hatching rate of zebrafish embryo was calculated after treated with various concentration synthesized ZnO NPs.

Developmental toxicity in embryos and larvae

To investigate the effect of CNPS on inducing the morphological changes it was treated to the zebrafish embryos. When different concentration of CNPs were exposed to the zebrafish embryos. The concentration of 10 µg/mL and 50 µg/mL showed the similar morphological level compared to the control. But in the group of higher concentration of 100 µg/mL, the changes in the morphology of zebrafish embryo was observed (Figure. 4). Already the higher concentration of 100 µg/mL showed toxic effect confirmed through previous experiments. These results suggest that the concentration of 10 µg/mL and 50 µg/mL are non toxic and safe to use.

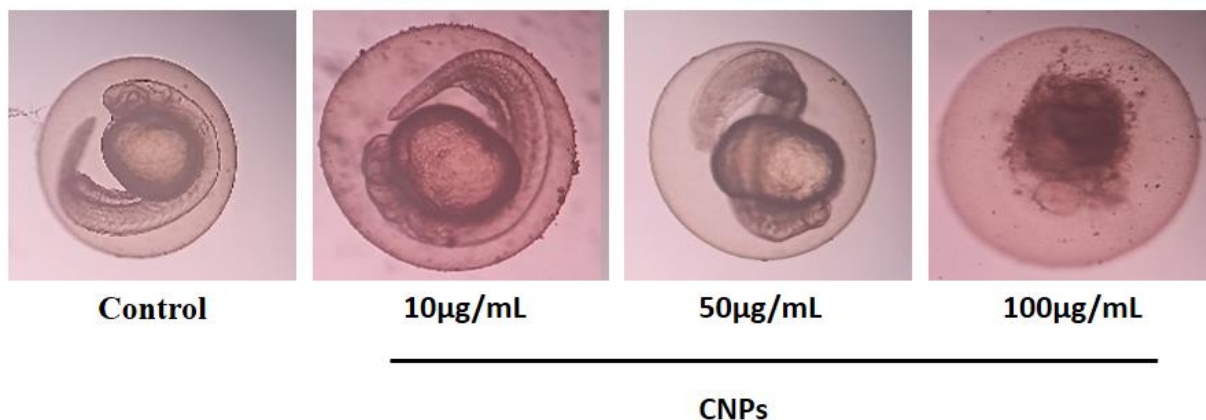


Fig 4: The images of zebrafish embryo after treated with CNPs.

Discussion:

In endodontics, chitosan nanoparticles (CNPs) are considered for various applications such as irrigants, intracanal medications, or root canal sealers due to their smaller size and compact structure. Their increased surface area allows for better absorption of medications and enhanced penetration into dentinal tubules [19-20]. While previous studies noted the efficacy of CNPs in reducing *E. faecalis* biofilm, their concentration and interaction time played crucial roles [21, 22].



Concerns about cytotoxicity and environmental hazards associated with CNPs have led to exploring green synthesis using plant extracts rich in phytochemicals, such as *Azadirachta indica* (*A. indica*) bark and *Semecarpus xanthocarpus* (*S. xanthocarpum*) seeds [23-25].

Zebrafish embryos, recognized as an excellent model for toxicology studies [26], were exposed to ZnO nanoparticles synthesized from *A. indica* leaves and *S. xanthocarpum* seed extracts. The study evaluated the toxicity in terms of survival rate, hatchability, and heart rate, revealing dose-dependent cytotoxicity. Previous reports indicated a correlation between CNP concentration reduction and toxicity reduction [27-29]. Comparisons with TiO₂ NPs suggested similar toxicity profiles due to their similar band gaps [30].

The study exposed zebrafish embryos to different concentrations of green-synthesized CNPs (10 µg/mL, 50 µg/mL, and 100 µg/mL). Results indicated alterations in hatching rates, with significant delays at higher concentrations (100 µg/mL). The study confirmed dose-dependent inhibitory effects on embryo hatching, aligning with previous reports [31-32]. Toxicity levels correlated directly with nanoparticle concentrations, reaching maximum toxicity at 100 µg/mL, resulting in a notable decrease in survival rates. Lower concentrations (10 µg/mL) showed no significant changes in survival rates, consistent with control embryos.

Observations of developmental defects and increased mortality at higher CNP concentrations were consistent with existing literature [33-36]. Morphological abnormalities, such as tail deformities and spinal curvature, were reported at the highest concentrations [37]. Heart rate reduction at higher concentrations could affect blood flow, potentially leading to depleted muscle glucose and lower muscular response, impacting hatching rates [38, 39].

Despite these findings, the green-synthesized CNPs showed no significant toxicities at low concentrations. However, the study suggests the need for further experiments, including long-term exposure studies *in vitro* and *in vivo*, to assess potential chronic effects and identify other influencing factors on CNP toxicity.

Conclusion:

Based on the results of this study, it can be concluded that Chitosan nanoparticles mixed with herbal extracts, at optimal concentrations, exhibit minimal developmental malformations and cytotoxic effects. This suggests their potential as intracanal medicaments in endodontic applications, emphasizing the importance of optimizing the dosage to ensure safety and effectiveness.

Limitations:

All these experiments were done under *in vitro* condition, clinical trials need to be conducted

Acknowledgments:

The authors would like to acknowledge [Funding Agency] for their financial support and [Your Institution] for providing the necessary facilities for this research.

Conflict of Interest: The authors declare no conflicts of interest

References:

Ajay Guru, Gokul Sudhakaran, S. Karthick Raja Namasivayam, Boopathi Seenivasan, Mukesh



- Pasupulieti, Jesu Arockiaraj, Meivelu Moovendhan, 2023. Serine Threonine-Protein Kinase-Derived IW13 Improves Lipid Metabolism via C/EBP- α /SREBP1/FAS Signaling Pathways in HFD-Induced Zebrafish In Vivo Larval Model. *Appl. Biochem. Biotechnol.* <https://doi.org/10.1007/s12010-023-04480-3>
- B, H.D.M., Guru, A., Sudhakaran, G., Murugan, R., Arshad, A., Arockiaraj, J., 2022. Double-edged sword role of shrimp miRNA explains an evolutionary language between shrimp-pathogen interactions that unties the knot of shrimp infection. *Rev. Aquac.* 14, 578–593. <https://doi.org/10.1111/raq.12613>
- Gopinath, P., Jesu, A., Manjunathan, T., Ajay, G., 2021. 6-Gingerol and semisynthetic 6-Gingerdione counteract oxidative stress induced by ROS in zebrafish. *Chem. Biodivers.* 11, 807–813. <https://doi.org/10.1002/cbdv.202100650>
- Guru, A., Arockiaraj, J., 2023. Exposure to environmental pollutant bisphenol A causes oxidative damage and lipid accumulation in Zebrafish larvae: Protective role of WL15 peptide derived from cysteine and glycine-rich protein 2. *J. Biochem. Mol. Toxicol.* 37. <https://doi.org/10.1002/jbt.23223>
- Guru, A., Issac, P.K., Saraswathi, N.T., Seshadri, V.D., Gabr, G.A., Arockiaraj, J., 2021a. Deteriorating insulin resistance due to WL15 peptide from cysteine and glycine-rich protein 2 in high glucose-induced rat skeletal muscle L6 cells. *Cell Biol. Int.* 45, 1698–1709. <https://doi.org/10.1002/cbin.11608>
- Guru, A., Issac, P.K., Velayutham, M., Saraswathi, N.T., Arshad, A., Arockiaraj, J., 2021b. Molecular mechanism of down-regulating adipogenic transcription factors in 3T3-L1 adipocyte cells by bioactive anti-adipogenic compounds. *Mol. Biol. Rep.* 48, 743–761. <https://doi.org/10.1007/s11033-020-06036-8>
- Guru, A., Lite, C., Freddy, A.J., Kumar, P., Pasupuleti, M., Saraswathi, T., Valan, M., Al-dhabi, N.A., Arshad, A., 2021c. Intracellular ROS scavenging and antioxidant regulation of WL15 from cysteine and glycine-rich protein 2 demonstrated in zebrafish in vivo model. *Dev. Comp. Immunol.* 114, 103863. <https://doi.org/10.1016/j.dci.2020.103863>
- Guru, A., Manjunathan, T., Sudhakaran, G., Juliet, A., Gopinath, P., Arockiaraj, J., 2023. 6-Gingerdione Reduces Apoptotic Conditions in HepG2 Cells and Inhibits Inflammatory Cytokine Gene Expression in Alcoholic Liver Injured Zebrafish Larvae. *Chem. Biodivers.* 20. <https://doi.org/10.1002/cbdv.202200959>
- Guru, A., Sudhakaran, G., Almutairi, M.H., Almutairi, B.O., Juliet, A., Arockiaraj, J., 2022a. β -cells regeneration by WL15 of cysteine and glycine-rich protein 2 which reduces alloxan induced β -cell dysfunction and oxidative stress through phosphoenolpyruvate carboxykinase and insulin pathway in zebrafish in-vivo larval model. *Mol. Biol. Rep.* <https://doi.org/10.1007/s11033-022-07882-4>
- Guru, A., Sudhakaran, G., Velayutham, M., Murugan, R., Pachaiappan, R., Mothana, R.A., Noman, O.M., Juliet, A., Arockiaraj, J., 2022b. Daidzein normalized gentamicin-induced nephrotoxicity and associated pro-inflammatory cytokines in MDCK and zebrafish: Possible mechanism of nephroprotection. *Comp. Biochem. Physiol. Part C Toxicol. Pharmacol.* 258,



109364. <https://doi.org/10.1016/j.cbpc.2022.109364>
- Haridevamuthu, B., Guru, A., Murugan, R., Sudhakaran, G., Pachaiappan, R., Almutairi, M.H., Almutairi, B.O., Juliet, A., Arockiaraj, J., 2022a. Neuroprotective effect of Biochanin a against Bisphenol A-induced prenatal neurotoxicity in zebrafish by modulating oxidative stress and locomotory defects. *Neurosci. Lett.* 790, 136889. <https://doi.org/10.1016/j.neulet.2022.136889>
- Haridevamuthu, B., Manjunathan, T., Guru, A., Kumar, R.S., Rajagopal, R., Kuppusamy, P., Juliet, A., Gopinath, P., Arockiaraj, J., 2022b. Hydroxyl containing benzo[b]thiophene analogs mitigates the acrylamide induced oxidative stress in the zebrafish larvae by stabilizing the glutathione redox cycle. *Life Sci.* 298, 120507. <https://doi.org/10.1016/j.lfs.2022.120507>
- Haridevamuthu, B., Manjunathan, T., Guru, A., Ranjith Wilson Alphonse, C., Boopathi, S., Murugan, R., Gatasheh, M.K., Atef Hatamleh, A., Juliet, A., Gopinath, P., Arockiaraj, J., 2022c. Amelioration of acrylamide induced neurotoxicity by benzo[b]thiophene analogs via glutathione redox dynamics in zebrafish larvae. *Brain Res.* 1788, 147941. <https://doi.org/10.1016/j.brainres.2022.147941>
- Issac, P.K., Guru, A., Velayutham, M., Pachaiappan, R., Arasu, M.V., Al-Dhabi, N.A., Choi, K.C., Harikrishnan, R., Arockiaraj, J., 2021a. Oxidative stress induced antioxidant and neurotoxicity demonstrated in vivo zebrafish embryo or larval model and their normalization due to morin showing therapeutic implications. *Life Sci.* 283, 119864. <https://doi.org/10.1016/j.lfs.2021.119864>
- Issac, P.K., Karan, R., Guru, A., Pachaiappan, R., Arasu, M.V., Al-Dhabi, N.A., Choi, K.C., Harikrishnan, R., Raj, J.A., 2021b. Insulin signaling pathway assessment by enhancing antioxidant activity due to morin using in vitro rat skeletal muscle L6 myotubes cells. *Mol. Biol. Rep.* 48, 5857–5872. <https://doi.org/10.1007/s11033-021-06580-x>
- Issac, P.K., Lite, C., Guru, A., Velayutham, M., 2021c. Tryptophan-tagged peptide from serine threonine-protein kinase of *Channa striatus* improves antioxidant defence in L6 myotubes and attenuates caspase 3 – dependent apoptotic response in zebrafish larvae National Centre for Cell Science. *Fish Physiol. Biochem.* <https://doi.org/10.1007/s10695-020-00912-7>
- Issac, P.K., Velayutham, M., Guru, A., Sudhakaran, G., Pachaiappan, R., Arockiaraj, J., 2022. Protective effect of morin by targeting mitochondrial reactive oxygen species induced by hydrogen peroxide demonstrated at a molecular level in MDCK epithelial cells. *Mol. Biol. Rep.* 1–12. <https://doi.org/10.1007/s11033-022-07261-z>
- Kathiravan, A., Manjunathan, T., Velusamy, M., Guru, A., Arockiaraj, J., Jhonsi, M.A., Gopinath, P., 2023. Nano-sized aggregation induced emissive probe for highly sensitive hypochlorous acid detection. *Dye. Pigment.* 210, 111016. <https://doi.org/10.1016/j.dyepig.2022.111016>
- Kumar, P., Ajay, I., Sri, G., Chandrakumar, S., Lite, C., 2020. Molecular process of glucose uptake and glycogen storage due to hamamelitannin via insulin signalling cascade in glucose metabolism. *Mol. Biol. Rep.* 47, 6727–6740. <https://doi.org/10.1007/s11033-020-05728-5>



- Lite, C., Guru, A., Juliet, M.J., Arockiaraj, J., 2022. Embryonic exposure to butylparaben and propylparaben induced developmental toxicity and triggered anxiety-like neurobehavioral response associated with oxidative stress and apoptosis in the head of zebrafish larvae. *Environ. Toxicol.* <https://doi.org/10.1002/tox.23545>
- Murugan, R., Guru, A., Haridevamuthu, B., Sudhakaran, G., Arshad, A., Arockiaraj, J., 2022a. Lantibiotics: an antimicrobial asset in combating aquaculture diseases. *Aquac. Int.* 30, 2365–2387. <https://doi.org/10.1007/s10499-022-00908-5>
- Murugan, R., Rajesh, R., Guru, A., Haridevamuthu, B., Almutairi, B.O., Almutairi, M.H., Juliet, A., Renganayagi, S., Gopinath, P., Arockiaraj, J., 2022b. Deacetylepoxyzadiradione Derived from Epoxyzadiradione of Neem (*Azadirachta indica* A. Juss) Fruits Mitigates LPS-Induced Oxidative Stress and Inflammation in Zebrafish Larvae. *Chem. Biodivers.* 19. <https://doi.org/10.1002/cbdv.202200041>
- Murugan, R., Rajesh, R., Seenivasan, B., Haridevamuthu, B., Sudhakaran, G., Guru, A., Rajagopal, R., Kuppusamy, P., Juliet, A., Gopinath, P., Arockiaraj, J., 2022c. Withaferin A targets the membrane of *Pseudomonas aeruginosa* and mitigates the inflammation in zebrafish larvae; an in vitro and in vivo approach. *Microb. Pathog.* 172, 105778. <https://doi.org/10.1016/j.micpath.2022.105778>
- Prabha, N., Guru, A., Harikrishnan, R., Gatasheh, M.K., Hatamleh, A.A., Juliet, A., Arockiaraj, J., 2022. Neuroprotective and antioxidant capability of RW20 peptide from histone acetyltransferases caused by oxidative stress-induced neurotoxicity in in vivo zebrafish larval model. *J. King Saud Univ. - Sci.* 100, 101861. <https://doi.org/10.1016/j.jksus.2022.101861>
- Priya, P.S., Guru, A., Meenatchi, R., Haridevamuthu, B., Velayutham, M., Seenivasan, B., Pachaiappan, R., Rajagopal, R., Kuppusamy, P., Juliet, A., Arockiaraj, J., 2023. Syringol, a wildfire residual methoxyphenol causes cytotoxicity and teratogenicity in zebrafish model. *Sci. Total Environ.* 864, 160968. <https://doi.org/10.1016/j.scitotenv.2022.160968>
- Sarkar, P., Guru, A., Raju, S. V., Farasani, A., Oyouni, A.A.A., Alzahrani, O.R., Althagafi, H.A.E., Alharthi, F., Karuppiyah, K.M., Arockiaraj, J., 2021. GP13, an *Arthrospira platensis* cysteine desulfurase-derived peptide, suppresses oxidative stress and reduces apoptosis in human leucocytes and zebrafish (*Danio rerio*) embryo via attenuated caspase-3 expression. *J. King Saud Univ. - Sci.* 33, 101665. <https://doi.org/10.1016/j.jksus.2021.101665>
- Siddhu, N.S.S., Guru, A., Satish Kumar, R.C., Almutairi, B.O., Almutairi, M.H., Juliet, A., Vijayakumar, T.M., Arockiaraj, J., 2022. Pro-inflammatory cytokine molecules from *Boswellia serrate* suppresses lipopolysaccharides induced inflammation demonstrated in an in-vivo zebrafish larval model. *Mol. Biol. Rep.* 7425–7435. <https://doi.org/10.1007/s11033-022-07544-5>
- Singh, M., Guru, A., Sudhakaran, G., Pachaiappan, R., Mahboob, S., Juliet, A., Gobi, M., Arockiaraj, J., 2022. Copper sulfate induced toxicological impact on in-vivo zebrafish larval model protected due to acacetin via anti-inflammatory and glutathione redox mechanism Comparative Biochemistry and Physiology , Part C Copper sulfate induced toxicological impact on i. *Comp. Biochem. Physiol. Part C* 262, 109463.



- <https://doi.org/10.1016/j.cbpc.2022.109463>
- Sudhakaran, G., Guru, A., Hari Deva Muthu, B., Murugan, R., Arshad, A., Arockiaraj, J., 2022a. Evidence-based hormonal, mutational, and endocrine-disrupting chemical-induced zebrafish as an alternative model to study PCOS condition similar to mammalian PCOS model. *Life Sci.* 291, 120276. <https://doi.org/10.1016/j.lfs.2021.120276>
- Sudhakaran, G., Guru, A., Haridevamuthu, B., Murugan, R., Arshad, A., Arockiaraj, J., 2022b. Molecular properties of postbiotics and their role in controlling aquaculture diseases. *Aquac. Res.* 53, 3257–3273. <https://doi.org/10.1111/are.15846>
- Sudhakaran, G., Prathap, P., Guru, A., Haridevamuthu, B., Murugan, R., Almutairi, B.O., Almutairi, M.H., Juliet, A., Gopinath, P., Arockiaraj, J., 2022c. Reverse pharmacology of Nimbin-N2 attenuates alcoholic liver injury and promotes the hepatoprotective dual role of improving lipid metabolism and downregulating the levels of inflammatory cytokines in zebrafish larval model. *Mol. Cell. Biochem.* 477, 2387–2401. <https://doi.org/10.1007/s11010-022-04448-7>
- Sudhakaran, G., Prathap, P., Guru, A., Rajesh, R., Sathish, S., Madhavan, T., Arasu, M. V., Al-Dhabi, N.A., Choi, K.C., Gopinath, P., Arockiaraj, J., 2022d. Anti-inflammatory role demonstrated both in vitro and in vivo models using nonsteroidal tetranortriterpenoid, Nimbin (N1) and its analogs (N2 and N3) that alleviate the domestication of alternative medicine. *Cell Biol. Int.* 46, 771–791. <https://doi.org/10.1002/cbin.11769>
- Sudhakaran, G., Rajesh, R., Guru, A., Arasu, M.V., Gopinath, P., Arockiaraj, J., 2023. Nimbin analogs N5 and N7 regulate the expression of lipid metabolic genes and inhibit lipid accumulation in high-fat diet-induced zebrafish larvae: An antihyperlipidemic study. *Tissue Cell* 80, 102000. <https://doi.org/10.1016/j.tice.2022.102000>
- Sudhakaran, G., Rajesh, R., Guru, A., Haridevamuthu, B., Murugan, R., 2022e. Deacetylated nimbin analog N2 fortifies alloxan-induced pancreatic β -cell damage in insulin-resistant zebrafish larvae by upregulating phosphoenolpyruvate carboxykinase (PEPCK) and insulin levels. *Toxicol. Appl. Pharmacol.* 454, 116229. <https://doi.org/10.1016/j.taap.2022.116229>
- Sudhakaran, G., Rajesh, R., Murugan, R., Velayutham, M., Guru, A., Boopathi, S., Muthupandian, S., Gopinath, P., Arockiaraj, J., 2022f. Nimbin analog N2 alleviates high testosterone induced oxidative stress in CHO cells and alters the expression of *Tox3* and *Dennd1a* signal transduction pathway involved in the PCOS zebrafish. *Phyther. Res.* 1–13. <https://doi.org/10.1002/ptr.7685>
- Velayutham, M., Guru, A., Arasu, M.V., Al-Dhabi, N.A., Choi, K.C., Elumalai, P., Harikrishnan, R., Arshad, A., Arockiaraj, J., 2021a. GR15 peptide of S-adenosylmethionine synthase (SAME) from *Arthrospira platensis* demonstrated antioxidant mechanism against H₂O₂ induced oxidative stress in in-vitro MDCK cells and in-vivo zebrafish larvae model. *J. Biotechnol.* 342, 79–91. <https://doi.org/10.1016/j.jbiotec.2021.10.010>
- Velayutham, M., Guru, A., Gatasheh, M.K., Hatamleh, A.A., Juliet, A., Arockiaraj, J., 2022a. Molecular Docking of SA11, RF13 and DI14 Peptides from Vacuolar Protein Sorting Associated Protein 26B Against Cancer Proteins and In vitro Investigation of its Anticancer



- Potency in Hep-2 Cells. *Int. J. Pept. Res. Ther.* 28. <https://doi.org/10.1007/s10989-022-10395-0>
- Velayutham, M., Ojha, B., Issac, P.K., Lite, C., Guru, A., Pasupuleti, M., Arasu, M.V., Al-Dhabi, N.A., Arockiaraj, J., 2021b. NV14 from serine O-acetyltransferase of cyanobacteria influences the antioxidant enzymes in vitro cells, gene expression against H₂O₂ and other responses in vivo zebrafish larval model. *Cell Biol. Int.* 45, 2331–2346. <https://doi.org/10.1002/cbin.11680>
- Velayutham, M., Sarkar, P., Sudhakaran, G., Al-Ghanim, K.A., Maboob, S., Juliet, A., Guru, A., Muthupandian, S., Arockiaraj, J., 2022b. Anti-Cancer and Anti-Inflammatory Activities of a Short Molecule, PS14 Derived from the Virulent Cellulose Binding Domain of *Aphanomyces invadans*, on Human Laryngeal Epithelial Cells and an In Vivo Zebrafish Embryo Model. *Molecules* 27, 7333. <https://doi.org/10.3390/molecules27217333>
- Choudhari, Sahil, Jogikalmat Krithikadatta, Ipsitha Vejendla, Swathi, and Mukesh Doble. 2023. “Microbial Interactions in Oral Biofilm: Evaluating Therapeutic Interventions and the Emergence of Resistance: A Narrative Review.” *Cureus*, October. <https://doi.org/10.7759/cureus.48021>.
- Deepika, Burra Anand, Jaiganesh Ramamurthy, Smiline Girija, and Nadathur Duraisamy Jayakumar. 2022. “Evaluation of the Antimicrobial Effect of *Azadiracta indica* L. Oral Gel against Anaerobic Oral Microbes: An in Vitro Study.” *World Journal of Dentistry* 13 (S1): S23–27.
- Janani, Krishnamachari, Kavalipurapu Venkata Teja, P. Ajitha, and Raghu Sandhya. 2020. “Evaluation of Tissue Inflammatory Response of Four Intracanal Medicament - An Animal Study.” *Journal of Conservative Dentistry: JCD* 23 (3): 216–20.
- Kamath, Ajith K., Iffat Nasim, N. P. Muralidharan, and Ravalika N. Kothuri. 2022. “Anti-Microbial Efficacy of Vanilla Planifolia Leaf Extract against Common Oral Micro-Biomes: A Comparative Study of Two Different Antibiotic Sensitivity Tests.” *Journal of Oral and Maxillofacial Pathology: JOMFP* 26 (3): 330–34.
- Lakshmi, and Dean -International Affairs, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences. 2021. “Green Synthesis of Copper Nanoparticles Synthesized Using Black Tea and Its Antibacterial Activity against Oral Pathogens.” *International Journal of Dentistry and Oral Science*, August, 4156–59.
- Lakshmi, Nanobiomedicine. n.d. “Department of Pharmacology, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Chennai.” *International Journal of Dentistry and Oral Science*, 2985–87.
- Nasim, Iffat, Zohra Jabin, S. Rajesh Kumar, and V. Vishnupriya. 2022. “Green Synthesis of Calcium Hydroxide-Coated Silver Nanoparticles Using *Andrographis paniculata* and *Azadiracta indica* Linn. Leaf Extracts: An Antimicrobial and Cytotoxic Activity.” *Journal of Conservative Dentistry: JCD* 25 (4): 369–74.

